

Biomimetic texturing as an effective antifouling solution



December 8th 2021

Chloe Richards, Adrián Delgado Ollero, Fiona Regan and Yan Delauré (Dublin City University)



Outline

- The problem of biofouling
- Traditional methods to prevent fouling
- Inspiration from nature
- Bioinspired design – the Brill fish, *Scophthalmus rhombus*
- Production methods
- Testing of designed micro-textures
- Results

The problem of biofouling



----->

Seconds to minutes Minutes to hours Hours to days Days to months Months to years



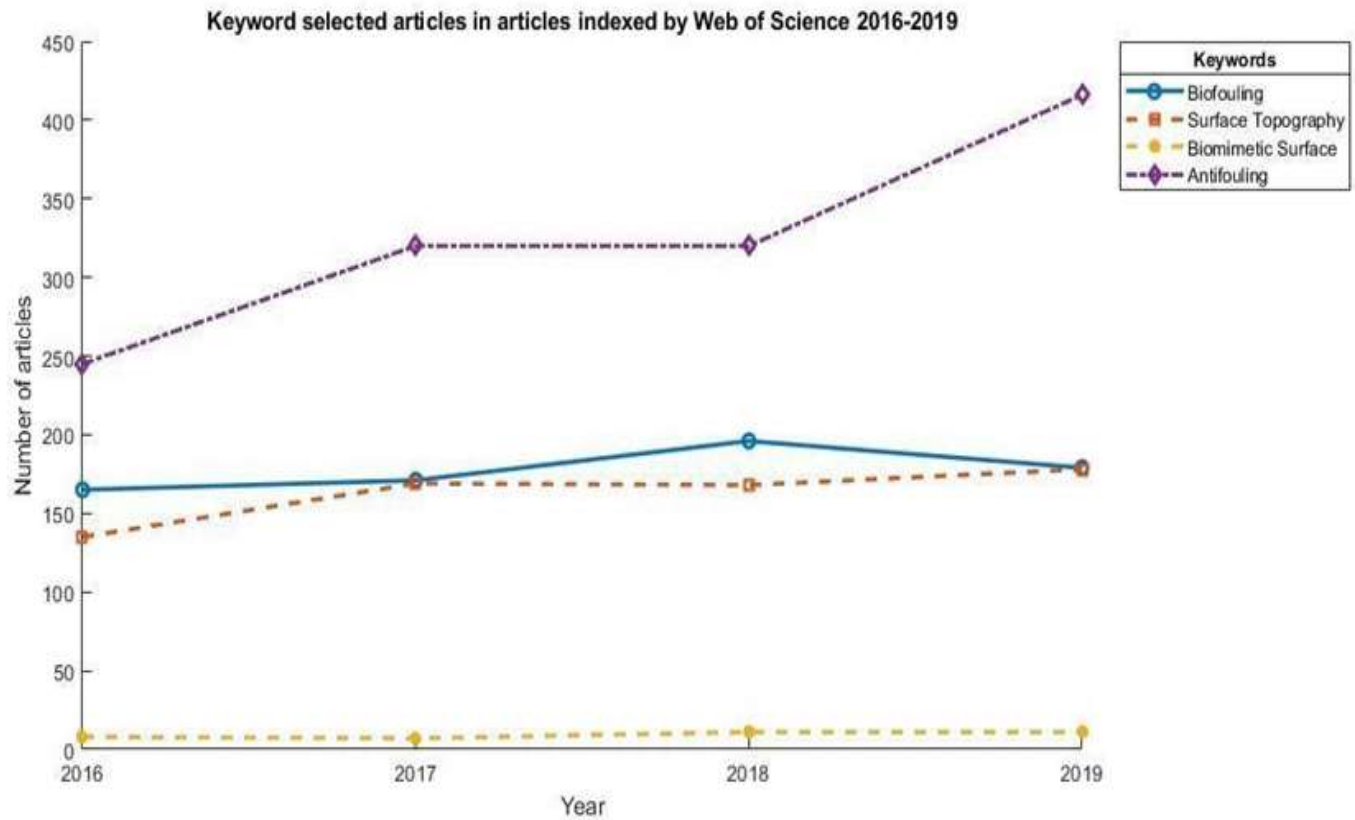
Traditional Methods

- Silicone Polymer
- Antifouling Paints
- Copper
- PVDF Resin
- Fouling Release Coatings
- TBT-SPC
- Fluorinated Ethylene Propylene
- Sol Gel Coatings
- Hydrophobic/Hydrophilic Coatings



Inspiration from nature

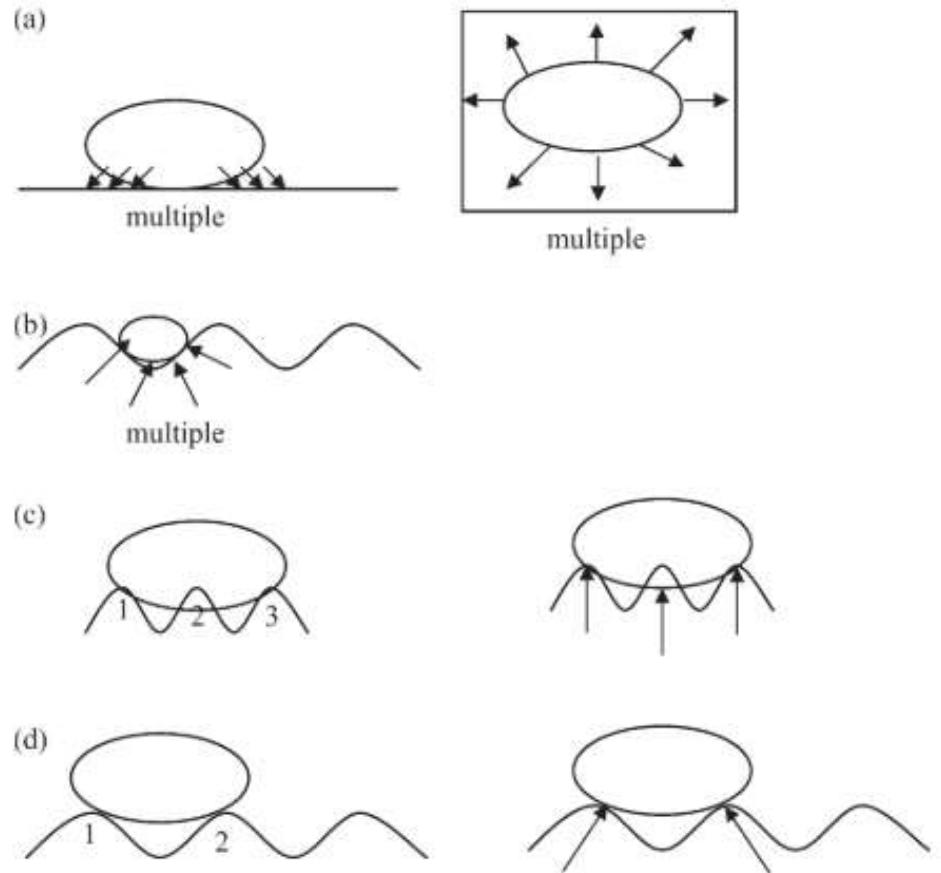




Int J Mol Sci. 2020 Jul 17;21(14):5063.



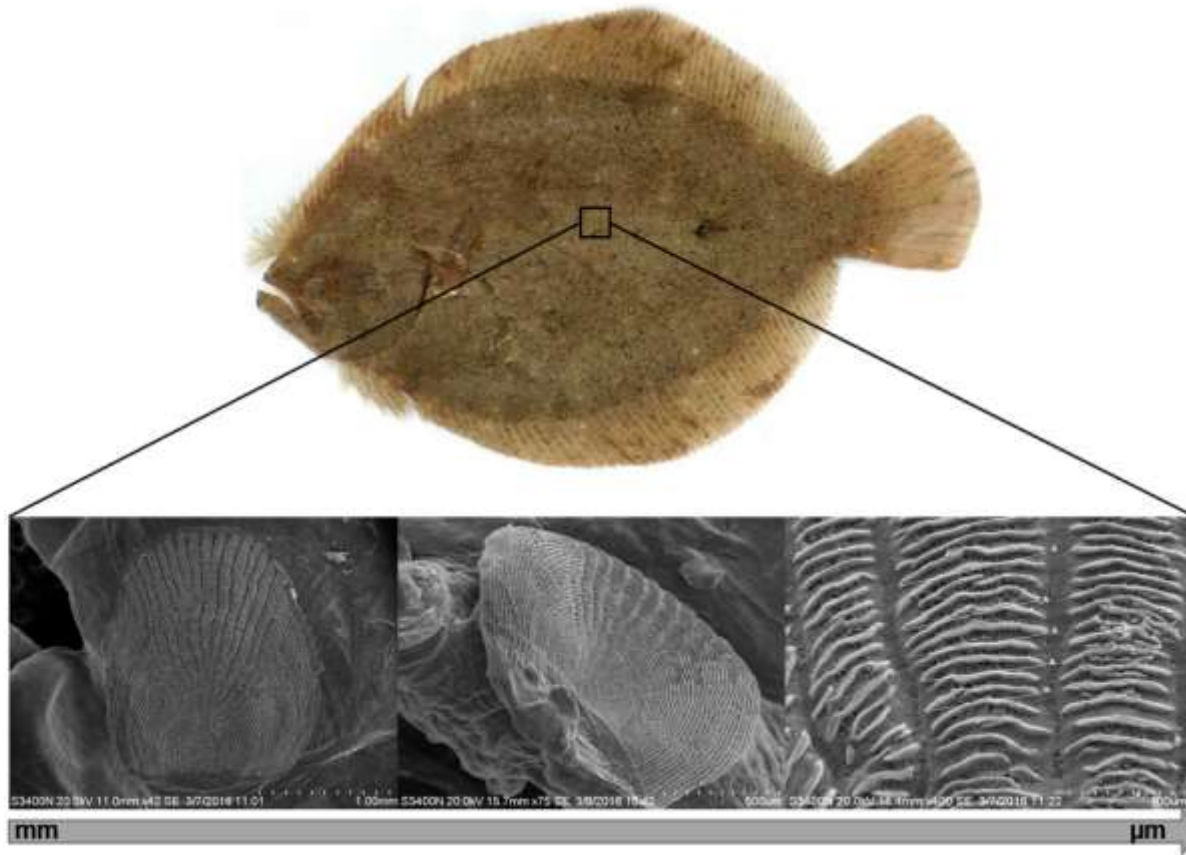
Why bioinspired surfaces?



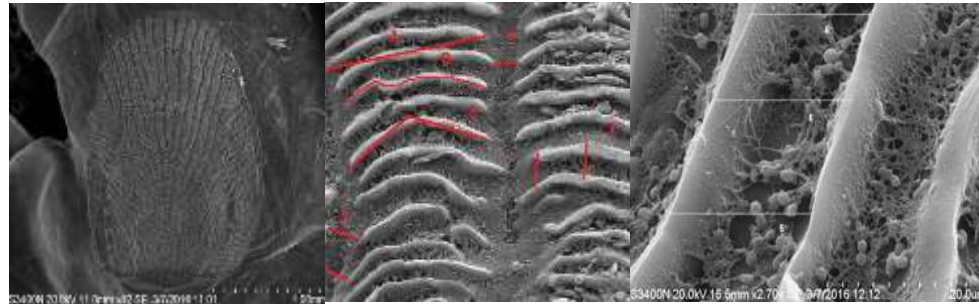
Biofouling, 2006; 22(1): 55 – 60



Marine design – the Brill fish (*Scophthalmus rhombus*)



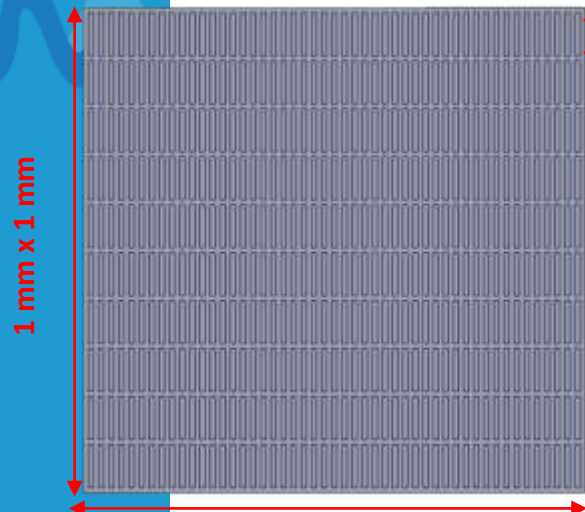
Production of bioinspired materials



- A: 74.84 μm
- B: 81.07 μm
- C: 143.08°
- D: 11.7 μm
- E: 16.6 μm
- F: 15.8 μm

1. Inspiration from nature, the Brill fish was characterized for antifouling potential.

2. Characterization of topography using SEM.



$L_x = 85 \mu\text{m}$

1 mm x 1 mm



$L_y = 10 \mu\text{m}$

$L_z = 10 \mu\text{m}$



3. Micro-texture design in SolidWorks 2017.

5. Production of micro-textures using 2-photon 3D printing.

3D Printing – Two photon polymerization (2PP)

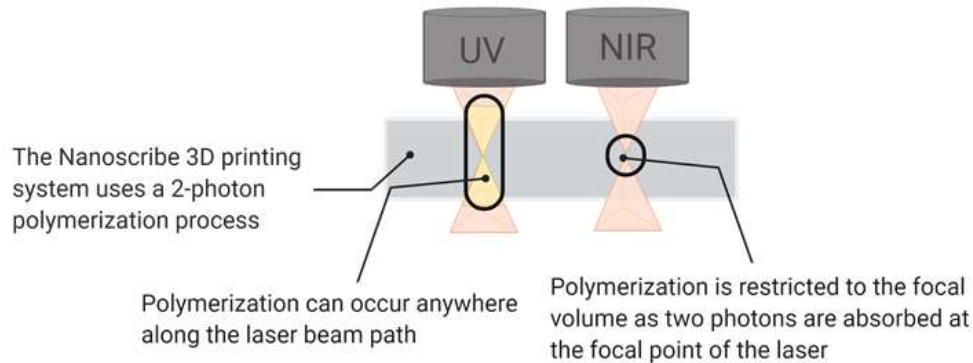


Figure 2. Nanoscribe 2-photon 3D printer



Figure 3. 3-D printed micro-texture taken using Keyence 3D light microscopy

Table 1. Summary of designed micro-textures

Label	Description
T1	Raised Bar Array (RBA) with sharpened edges and L_y dimension of $10\ \mu\text{m}$
T2	Raised Bar Array (RBA) with rounded edges and L_y dimension of $10\ \mu\text{m}$

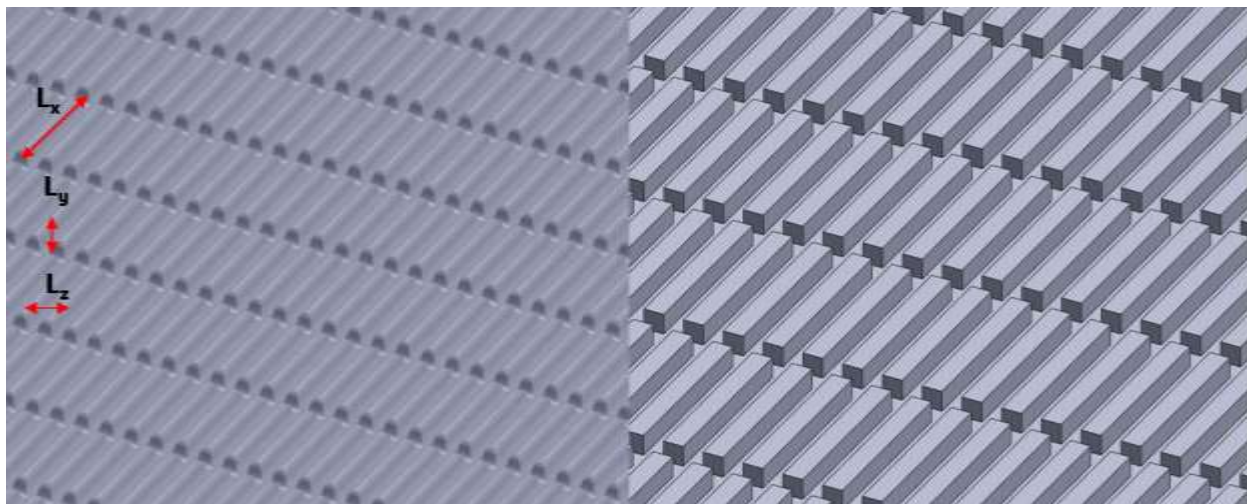


Figure 1. Outline sketch of Raised Bar Array (RBA) micro-texture for feature height $L_y = 10\ \mu\text{m}$ and $L_z = 10\ \mu\text{m}$

Testing of designed micro-textures

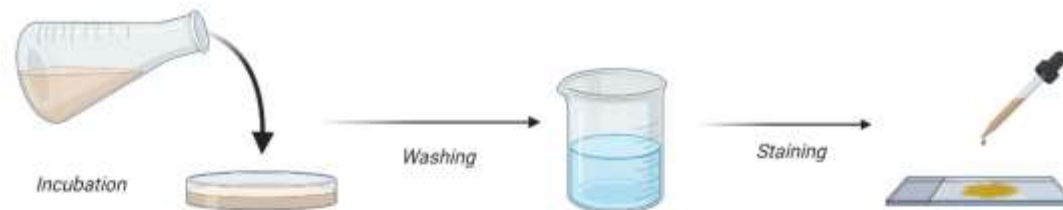
Step 1

Cell numbers were estimated by counting in the central area of an improved Neubauer hemocytometer.



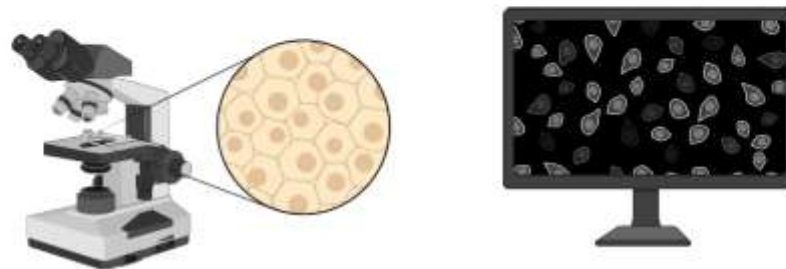
Step 2

Microtextures were immersed in 10 mL culture suspension for 3 hr.

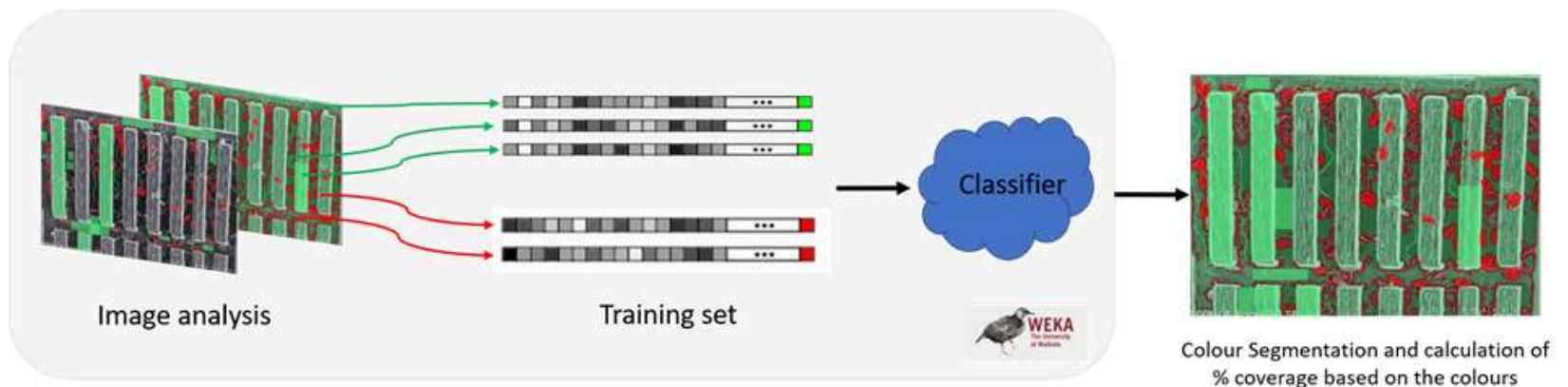
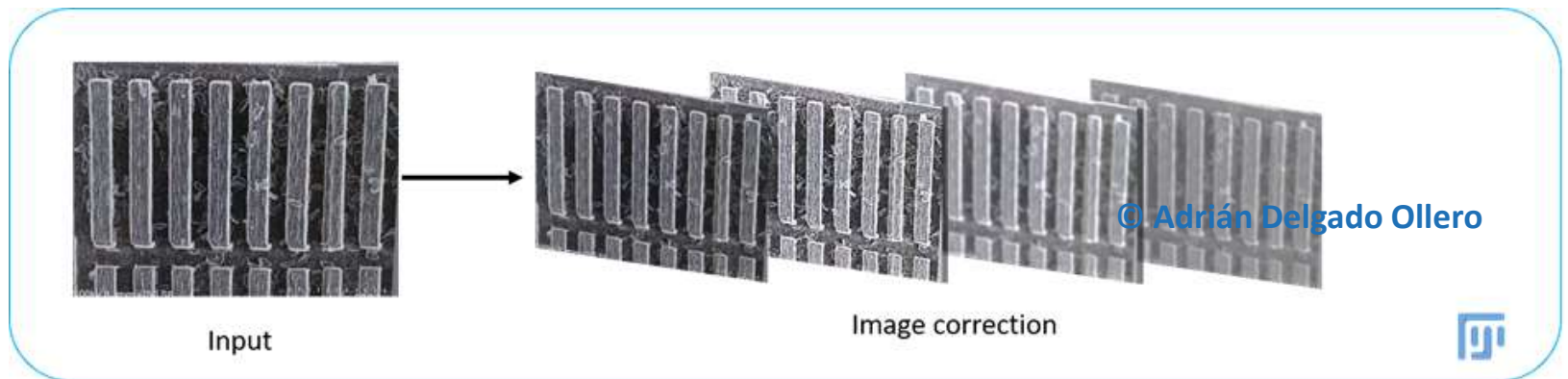


Step 3

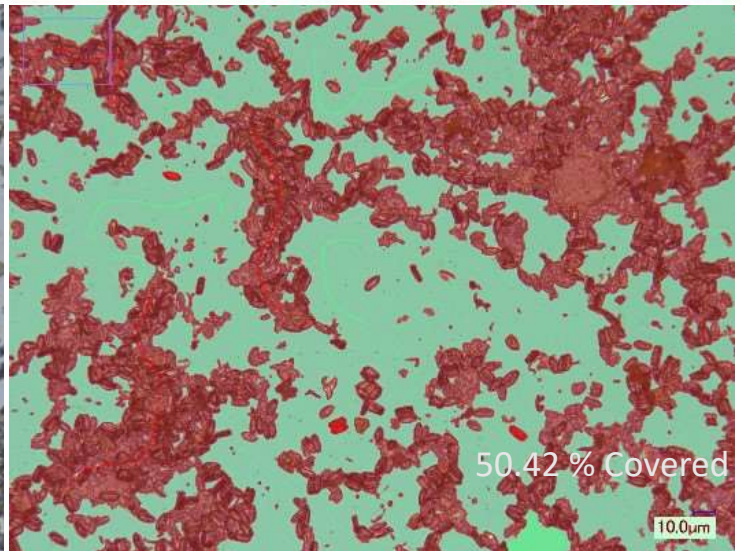
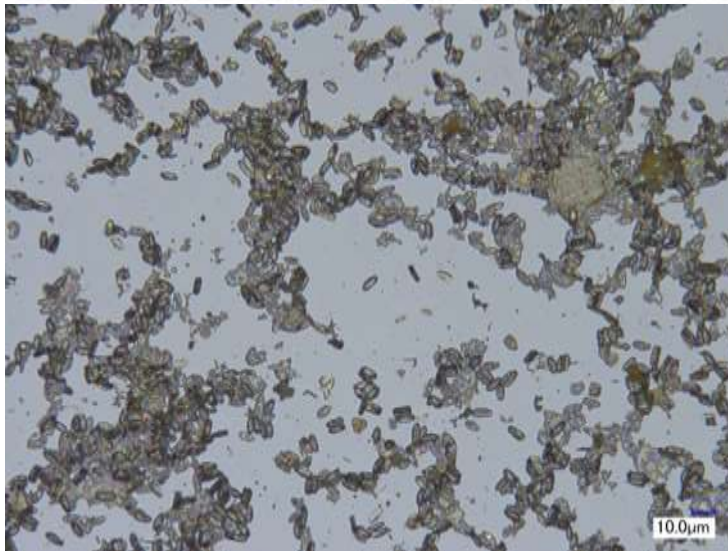
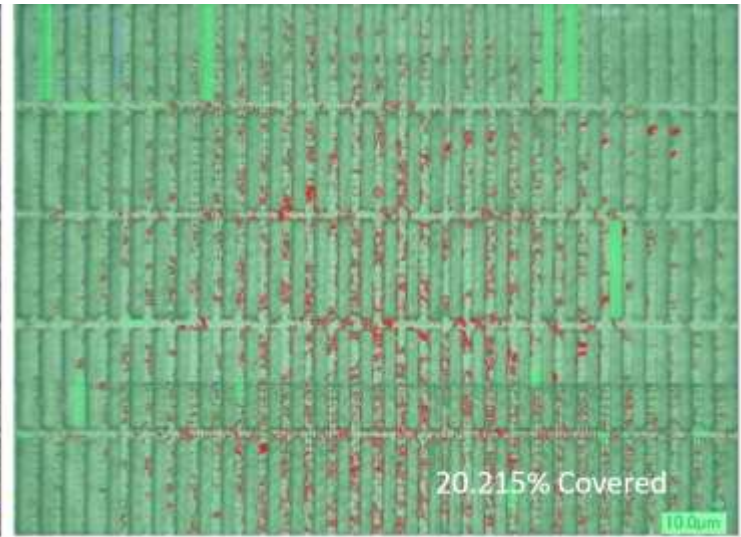
Colonization of microtextures was observed using light microscopy and ImageJ image processing software.



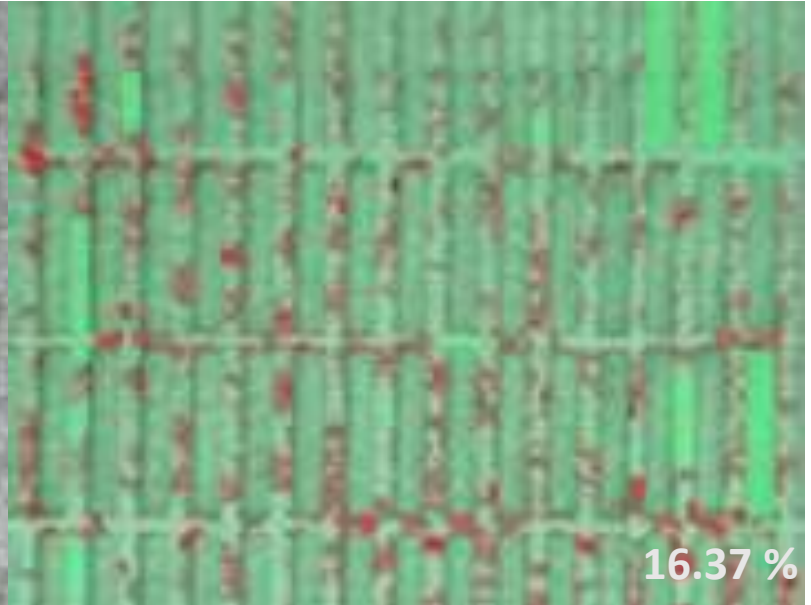
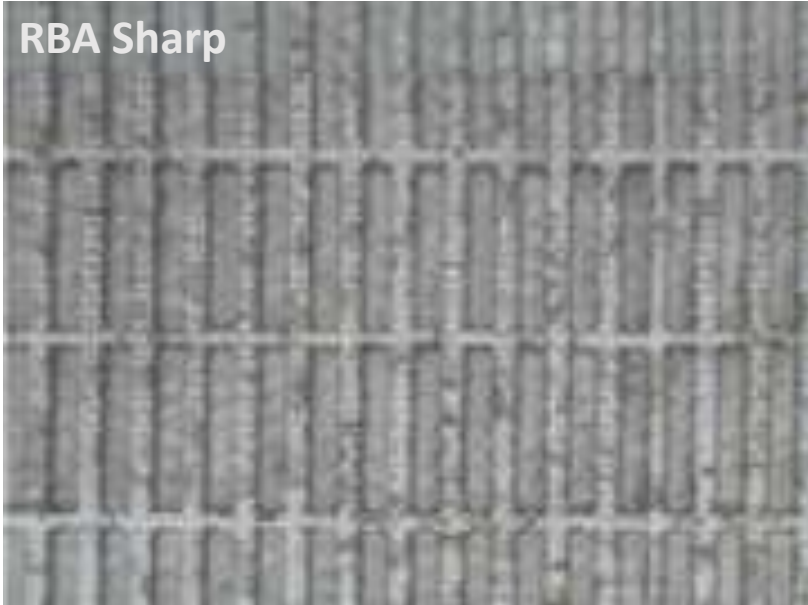
Statistical analysis using ImageJ



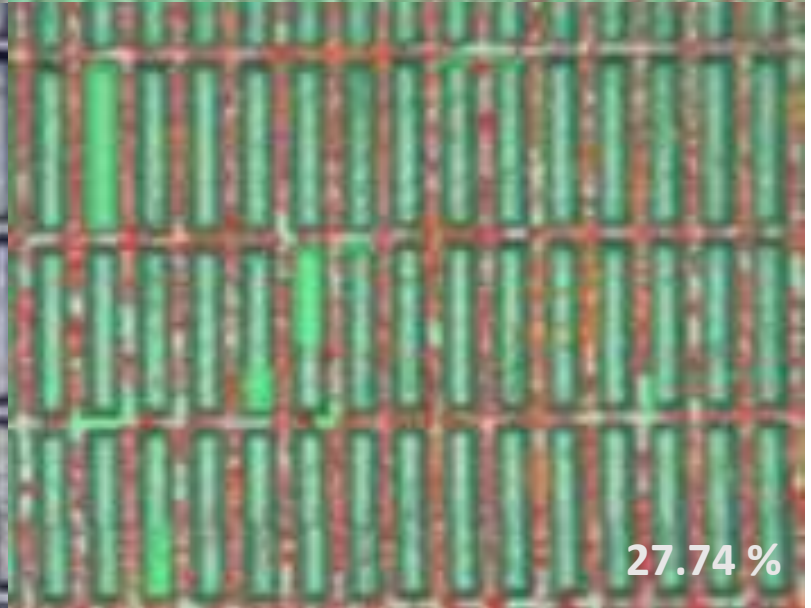
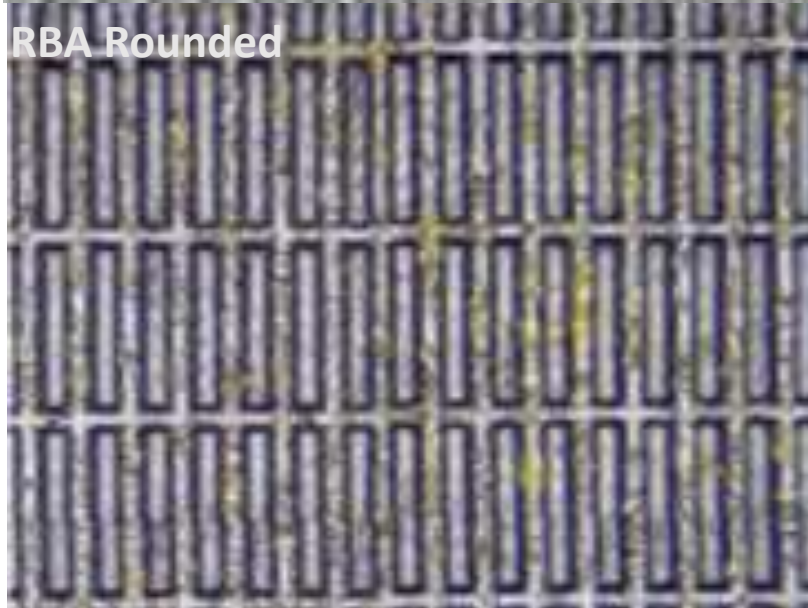
Results



RBA Sharp



RBA Rounded



Results

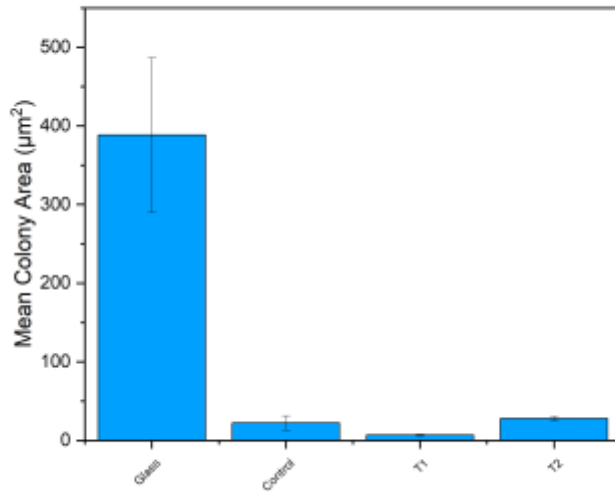


Figure 2. Mean colony area (μm^2) of *N. ovalis* cells on produced textured surfaces. Standard error was used to produce error bars ($n = 3$)

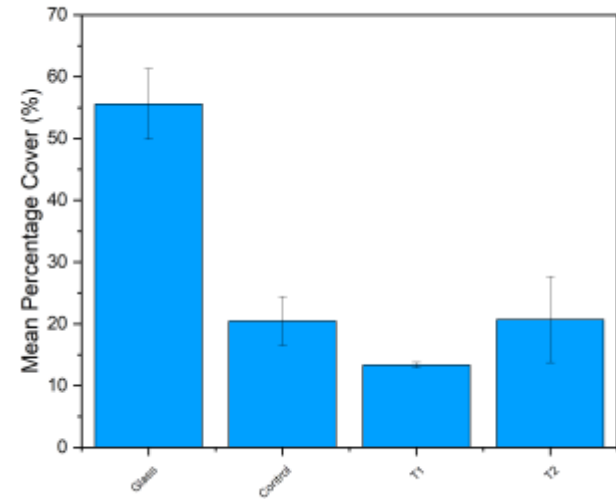


Figure 3. Mean biofilm cover (%) of *N. ovalis* cells on produced textured surfaces ($n = 3$)

Acknowledgements

Prof Fiona Regan

Dr Yan Delauré

Adrián Delgado Ollero

Philip Daly

Dr Ciprian Briciu-Burghina

Alan Barrett

Dr Ivan Maguire

Funding & Technical Support

- EU Horizon 2020 Research & Innovation Programme
- Marine Institute
- DCU Water Institute
- School of Chemical Sciences
- Chemistry Technical Team
- NRF Technical Team



Thank you for your attention!

Chloe Richards

chloe.richards3@mail.dcu.ie

www.nemmo.eu



[@Nemmo_Project](https://twitter.com/Nemmo_Project)



info@nemmo.eu



This project has received funding from the European Union's Horizon 2020 research and innovation programme under grant agreement No 815278.

